When binding affinity matters.

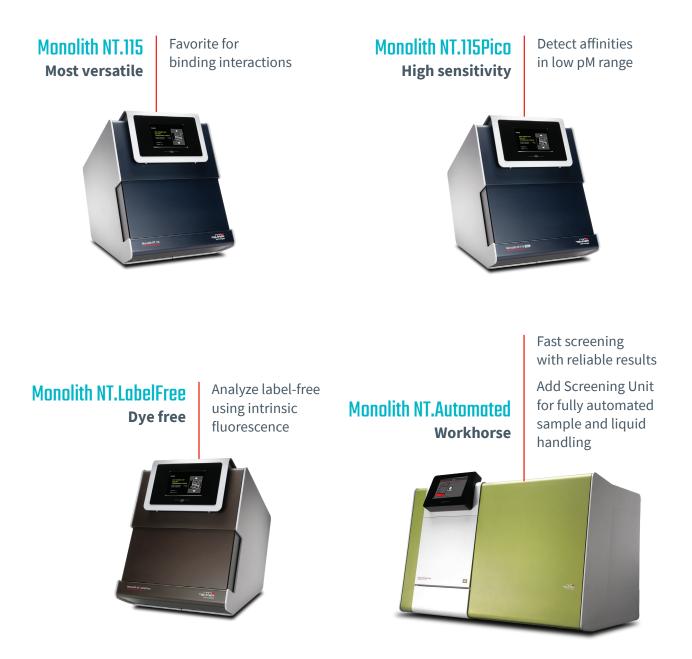
Swiftly measure binding interactions using very little sample with the Monolith





When choices matter.

Anyone can measure binding affinity in minutes. Each system is simple to use and not only has a friendly lab footprint but is maintenance-free.



When decisions matter.

Move forward with the best candidates quickly—get accurate K_d values in minutes using very little sample consumption.

Get results quickly

Capture measurements in minutes not hours or even days.

Measure any sample type

Easily measure interactions between molecules of different molecular weights or classes.

Consume very little sample

Determine dissociation constants using a lot less sample than other methods require.

Do minimal sample prep

Analyze samples in any buffer or bioliquid and in close-to-native conditions without purification or immobilization steps.

Optimize assays easily

Feedback is immediate making optimization of assays so much easier.

Evaluate the most difficult targets

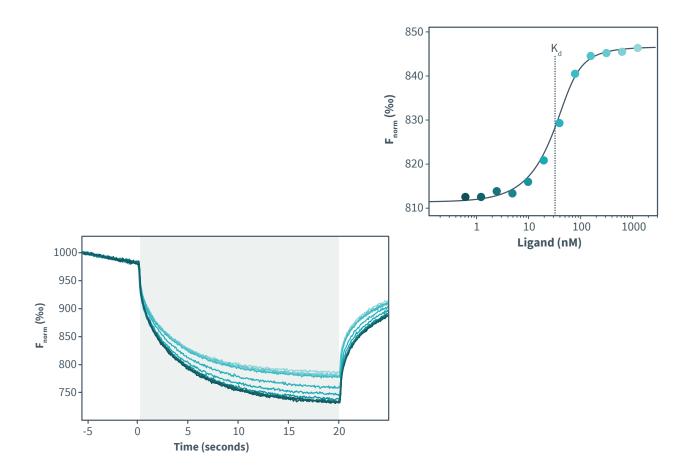
Running into a roadblock? Get results even when traditional methods don't work.

When solutions matter.

Monolith uses MST technology to quantify binding events. One partner is typically labeled with a fluorescent dye and becomes an extremely sensitive reporter for all binding interactions. These events, weak or strong, cause a change in MST signal that is easily detected. The results are automatically translated into precise and quantitative K_d values.

Getting accurate results quickly requires the use of high quality capillaries. Advantages are numerous — minute amounts of samples are analyzed in-solution, under native or close-to-native conditions, and in an immobilization-free environment without the need for purification.





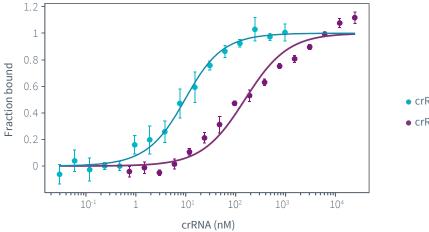
When confidence matters.

Monolith is all about accurately determining binding affinity in minutes. The strength of the interactions between a fluorescently labeled or intrinsically fluorescent sample and a binding partner (or ligand) are measured while a temperature gradient is applied (gray box) over time (left figure). From this, binding affinity (K_d) is automatically calculated from a fitted curve that plots normalized fluorescence against concentration of ligand (right figure). Get answers for 1, 16 or even 96 samples at a time, and be confident that results are precise and complete.

When insight matters.

More knowledge means more decision-making power. Determining binding affinity between large or small molecules, nucleic acids or even ions provides answers to important mechanisms of action or regulatory pathways.

For binding affinities, the lower the K_d, the higher the strength of the binding interaction between two molecules. By understanding this with the Monolith, Dong et al. finally had a view into how cellular expression and regulation work in their CRISPR RNA model.



Monitoring MG2+ dependence of CRISPR RNA to a Cas9-like nuclease (Dong et al., Nature, 532:522-536 2016)



• crRNA (10mM EDTA), K_d = 154.78 ± 36.73 nM

Session	13 🐼 Binding Affinity 🔹 🕦 Plan					nstructions 3 Results 4 Details Temperature Contro				ontrol Off		
Overview						MST Buffer including 0.05% Tw	🔆 Auto-de	tect Red			You can	enter here so
E Save					0	Unknown	₩ Medium					
Save	Plan Your Experiment											A Bind
good	K Target					🕒 Ligand						quantil purpos observ
linding Check orescence	MyTarget		~	?]	MyLigand				~	?	of liga ligand
1., adsorption, gregation	Use His-Tag Labeling			?]	Estimated Kd		optional	μΜ	~	?	are ad Measu extract
Sinding Affinity good	Concentration of stock solution		μΜ ~	?		Concentration of stock solut	ion		μΜ	~	?	For a
Sinding Affinity	Concentration in this assay	20nM	1	?		Ligand in organic solvent lik	e DMSO				?	to perf
t good (aggr.) Jinding Affinity	Assay buffer					Ligand buffer in this assay		-			?	
t good (fluor.)	MST Buffer including 0.05% Tween		1	?]	Highest concentration in this	s assay	-	1	**	?	
Sinding Affinity Iorescence ., aggregation	Capillary											
Sinding Affinity			~	?								
o binding	System settings											
Sinding Affinity I. fluor. change	Excitation Power	MST-Powe	r									
iD-Test ecificity Test	Auto-detect 20 % 🗘 ? M	edium 🜱	1	?								

When answers matter.

Anyone in the lab can run binding interaction experiments — MO.Control software calculates all the volumes and dilutions needed to run an assay. With guided setup, real-time optimization tips, and built-in knowledge-based advice, users feel confident their experiments will run smoothly. MO.Control saves time by automatically calculating binding affinity values and instantly generating complete result summaries. Exporting data and reporting results is super easy with just one click.

When fit matters.

	NT.115	NT.115 Pico	NT. LabelFree	NT.Automated				
Experimental time	15 minutes or less							
Dynamic range	1 nM to mM	1 pM to mM	10 nM to mM	1 pM to mM				
Detected molecule range								
Samples per run		16 manually		96 manually, higher throughput with NT.Automated Screening Unit				
Minimum sample volume measured		4 µL		3 μL				
Temperature control	7/(10)							
Fluorescence channels	1 (Pico-Red)		1 (UV)	Up to 4 (Blue, Green, Red, Pico-Red, UV)				
Dimensions	33 (85 cm W x 63 cm H x 57 cm D						
Weight		70 kg						
Optional upgrade				NT.Automated Screening Unit (for automated sample and liquid handling)				

A selection of assay kits, control kits, labeling kits as well as specific capillary types are available to measure binding affinities for many protein types. Visit nanotempertech.com to see the entire list of consumables.



nanotempertech.com